Salmon River Hatchery

INTRODUCTION

Salmon River Hatchery is located seven miles north of Lincoln City off Highway 18 near Otis. Site is at an elevation of 30 feet above sea level, at latitude 45.0172 and longitude -123.9364. Site area is 23.67 acres.

The hatchery water supply is provided by five electric pumps on Salmon River. Water rights are for 30.03 cfs.

The facility is staffed with 3 FTE.

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PURPOSE

Salmon River Hatchery was constructed in 1975 and facility operations are funded by the State of Oregon.

The facility is used for adult collection, egg incubation and rearing of fall Chinook and egg incubation and rearing of Rainbow Trout and Coho.

PROGRAM TYPE

The ODFW Hatchery Management Policy defines hatchery programs as either harvest or conservation programs. Harvest programs operate to enhance or maintain fisheries without impairing naturally reproducing populations. Conservation programs operate to maintain or increase the number of naturally produced fish without reducing the productivity of naturally reproducing populations.
Salmon River Hatchery programs are harvest programs, used for the augmentation of fishing and harvest opportunities.

GOALS

Fall Chinook:

Salmon River (36H) stock: to provide hatchery-produced fish for sport and commercial harvest in both the ocean environment and the Salmon River. This stock serves as one of the indicator stocks for the US/Canada treaty.

Coho:

Big Creek (13H) stock: to mitigate for reduced coho salmon catch in sport and commercial fisheries due to habitat and passage loss or degradation in the Columbia River Basin by providing a high quality coho salmon for harvest in Lower Columbia River (LCR) Select Area commercial and recreational fisheries.

Rainbow Trout:

Cape Cod Triploid (72T) stock: to produce legal-size and trophy triploid rainbow trout to augment the trout fishery in coastal lakes by providing catchable hatchery trout in the spring.

OBJECTIVES

Objective 1: Foster and sustain opportunities for sport, commercial, and tribal fishers consistent with the conservation of naturally produced native fish.

Fall Chinook:

Salmon River (36H) stock
Produce 200,000 smolts (14,286 pounds) for release into the Salmon River.

Coho:

Big Creek (13H) stock
Produce 385,000 pre-smolts (12,833 pounds) for transfer to Clatsop County Fisheries South Fork Klaskanine facility.

Rainbow Trout:

Cape Cod Triploid (72T) stock
Produce 30,000 legals (15,000 pounds) and 4,500 trophies (4,500 pounds) for release into various waterbodies in the North Coast and Mid Coast districts.

Objective 2: Contribute toward the sustainability of naturally produced native fish populations through the responsible use of hatcheries and hatchery-produced fish.

Objective 3: Maintain genetic resources of native fish populations spawned or reared in captivity.

Objective 4: Restrict the introduction, amplification, or dissemination of disease agents in hatchery produced fish and in natural environments by controlling egg and fish movements and by prescribing a variety of preventative, therapeutic and disinfecting strategies to control the spread of disease agents in fish populations in the state.

Objective 5: Minimize adverse ecological impacts to watersheds caused by hatchery facilities and operations.
Objective 6: Communicate effectively with other fish producers, managers and the public.

CURRENT PRACTICES TO ACHIEVE OBJECTIVES

The sections that follow describe the current hatchery practices associated with fish production at this facility. Because ODFW hatcheries are managed to maximize use of the hatchery rearing space, hatchery operations are dynamic and subject to annual change depending upon statewide program needs.

The Native Fish Conservation Policy, the Fish Hatchery Management Policy, the Fish Health Management Policy and Hatchery Genetic Management Plans provide guidelines for the management of wild and hatchery fish in Oregon. These policies describe the brood collection, rearing, release, and health management strategies currently used at this facility.

Objective 1: Foster and sustain opportunities for sport, commercial, and tribal fishers consistent with the conservation of naturally produced native fish.

Adult Collection

Fall Chinook:
Salmon River (36H) stock: Adults arrive at the hatchery between September and November. Peak spawning occurs in November. The annual adult collection goal is 75 pairs.

Coho:
Big Creek (13H) stock: No adults are collected at this facility. Refer to the Big Creek Hatchery Plan for details.

Rainbow Trout:
Cape Cod Triploid (72T) stock: Broodstock are maintained at Roaring River Hatchery.

Objective 2: Contribute toward the sustainability of naturally produced native fish populations through the responsible use of hatcheries and hatchery-produced fish.

Rearing and Release Strategies

Rearing and release strategies are designed to limit the amount of ecological interactions occurring between hatchery and naturally produced fish. Fish are reared to sufficient size that smoltification occurs within nearly the entire population, which will reduce the retention time in downstream migration. Rearing on parent river water, or acclimation to parent river water for several weeks, is used to ensure strong homing to the hatchery, thus reducing the stray rate to natural populations. Various release strategies are used to ensure that fish migrate from the hatchery with least amount of interaction with native populations. The specific rearing and release strategies used at this hatchery are outlined below.
Fall Chinook:
  Salmon River (36H) stock: Produce 200,000 smolts at a size of 14 fpp for on-station release into the Salmon River at the optimal time between August 15th and September 15th. All fish are fin-clipped and coded-wire tagged prior to release.

Coho:
  Big Creek (13H) stock: Produce 385,000 pre-smolts at a size of 30 fpp for transfer to the CCF South Fork Klaskanine facility in early October. All fish are fin-clipped and 25,000 are coded-wire tagged prior to release.

Rainbow Trout:
  Cape Cod Triploid (72T) stock: Produce 30,000 legals at a size of 2 fpp for release into various waterbodies in March and April.
  Produce 4,500 trophy trout at a size of 1 fpp for release into various waterbodies in June.

Objective 3: Maintain genetic resources of native fish populations spawned or reared in captivity.

Broodstock Selection and Spawning

Oregon's Native Fish Conservation Policy and Hatchery Genetic Management Plans outline broodstock selection and spawning protocols for some fish stocks. The following practices are currently being used at Salmon River Hatchery:

Fall Chinook:
  Salmon River (36H) stock: Adults are collected at the Salmon River Hatchery trap throughout the run. The goal of the program is to use 50% wild adults in the broodstock each year. Broodstock is selected based on available ripe fish. Mating and spawning are done on a 1:1 sex ratio.

Coho:
  Big Creek (13H) stock: Broodstock selection and spawning take place at Big Creek Hatchery (see the Big Creek Hatchery Plan for details).

Rainbow Trout:
  Cape Cod Triploid (72T) stock: Broodstock selection and spawning take place at Roaring River Hatchery (see the Roaring River Hatchery Plan for details).

Objective 4: Restrict the introduction, amplification, or dissemination of disease agents in hatchery produced fish and in natural environments by controlling egg and fish movements and by prescribing a variety of preventative, therapeutic and disinfecting strategies to control the spread of disease agents in fish populations in the state.

Fish Health Management Programs--All Stocks

The primary objective of fish health management programs at ODFW hatcheries is to produce healthy smolts that will contribute to the fishery and return sufficient numbers of adults to continue propagation of the stocks and provide supplementation if desired. Equally important is to prevent the
introduction, amplification or spread of fish pathogens that might negatively affect the health of both hatchery and naturally reproducing stocks.

ODFW has implemented both disease control and disease prevention programs at all of its facilities to achieve these objectives. These programs include the following standard elements:

**Disease Control (Reactive)**
- Perform necropsies of diseased and dead fish to diagnose the cause of loss.
- Prescribe appropriate treatments and remedies to disease. This includes recommending modifications in fish culture practices, when appropriate, to alleviate disease-contributing factors.
- Apply a disease control policy as stated in the Oregon Administrative Rules which dictates how specific disease problems will be addressed and what restrictions may be placed on movements of diseased stocks.
- Conduct applied research on new and existing techniques to control disease epizootics.

**Disease Prevention (Proactive)**
- Routinely remove dead fish from each rearing container and notify ODFW Fish Pathology if losses are increasing. Monthly mortality records are submitted to Fish Pathology from each hatchery.
- Routinely perform examinations of live fish to assess health status and detect problems before they progress to clinical disease or mortality.
- Implement disease preventative strategies in all aspects of fish culture to produce a quality fish. This includes prescribing the optimal nutritional needs and environmental conditions in the hatchery rearing container based on historical disease events. It also involves the use of vaccines or antibiotics in order to avoid a disease problem.
- Use a disease prevention policy that restricts the introduction of stocks into a facility. This will help avoid new disease problems and fish pathogens not previously found at the site.
- Use sanitation procedures that prevent introduction of pathogens into and/or within a facility.
- Conduct applied research on new and existing disease prevention techniques.
- Utilize pond management strategies (e.g., Density Index and Flow Index guidelines) to help optimize the quality of the aquatic environment and minimize fish stress that can be conducive to infectious and noninfectious diseases. For example, a Density Index is used to estimate the maximum number of fish that can occupy a rearing unit based on the rearing unit’s size. A Flow Index is used to estimate the rearing unit’s carrying capacity based on water flows.

**Fish Health Activities at Salmon River Hatchery**

**Health Monitoring**
- All fish are given a health inspection no longer than 6 weeks before fish are released or transferred. This exam may be in conjunction with the routine monthly visit.
- Monthly health monitoring examinations of healthy and clinically diseased fish are conducted on each fish lot at the hatchery.
- Examinations for *Myxobolus cerebralis*, agent of whirling disease, are conducted annually on 60 fish held for a minimum of 180 days at the facility.
• At spawning, a minimum of 60 ovarian fluids and 60 kidney/spleen/pyloric caeca (based on a minimum sampling at the 5% incidence level) are examined for viral pathogens from each salmon lot. If prespawning mortality is above normal, necropsies are conducted on dead adult fish for bacteria, parasites and other causes of death.

• Whenever abnormal behavior is reported or observed, or mortality exceeds 0.1% per day over five consecutive days in any rearing container, the fish pathologist will examine the affected fish, make a diagnosis and recommend the appropriate remedial or preventative measures.

• Reporting and control of specific fish pathogens are conducted in accordance with the Fish Health Management Policy. Results from each examination mentioned above are reported on the ODFW Fish Health or Virus Examination forms.

Fish and Egg Movements

• Movements of fish and eggs are conducted in accordance with the Fish Health Management Policy.

Therapeutic and Prophylactic Treatments

• Juvenile fish are administered antibiotics orally as needed for the control of bacterial infections and for prevention of diseases.

• Formalin is dispensed into water for control of parasites and fungus on eggs. Treatment dosage and exposure time varies with species, life stage and condition being treated.

• Hydrogen Peroxide is dispensed into water for control of external parasites on juveniles. Treatment dosages and exposure time varies with species, life stage and condition being treated.

• Only approved or permitted therapeutic agents are used for treatments:
  o FDA labeled and approved for use on food fish
  o Allowed by the FDA as an Investigational New Animal Drug
  o Obtained by extra-label prescription from a veterinarian
  o Allowed by the FDA as low regulatory priority or deferred regulatory status
  o Approved by the FDA through USFWS for fish listed under the federal Endangered Species Act.

Sanitation

• All eggs brought to the facility are surface-disinfected or water-hardened in buffered iodophor.

• Disinfection footbaths (or other means of disinfection) are provided at the incubation facility’s entrance and exit areas while embryos are incubating in the facility.

• All equipment (e.g., nets, tanks, rain gear, boots) is disinfected with iodophor between uses with different fish/egg lots or different rearing containers.

• Dead fish are disposed of promptly and in a manner that prevents introduction of disease agents to the waters of the state.

• Rearing units are cleaned on a regular basis.

• Fish transport trucks are disinfected between the hauling of different fish lots. Rearing units are sanitized after removing fish and before introducing a new fish stock either by thorough cleaning and use of a disinfectant or by cleaning and leaving dry for an extended time.
Rearing units are sanitized after removing fish and before introducing a new fish stock either by thorough cleaning and use of a disinfectant or by cleaning and leaving dry for an extended time.

Objective 5: Minimize adverse ecological impacts to watersheds caused by hatchery facilities and operations.

Environmental Monitoring

Primarily, environmental monitoring is conducted at ODFW facilities to ensure these facilities meet the requirements of the National Pollution Discharge Elimination Permit administered by the Oregon Department of Environmental Quality. It is also used in managing fish health. On a short-term basis, monitoring helps identify when changes to hatchery practices are required. Long-term monitoring provides the ability to quantify water quality impacts resulting from changes in the watershed (e.g., logging, road building and urbanization). The following environmental parameters are currently monitored at all ODFW hatcheries:

- **Total Suspended Solids (TSS)** – measured quarterly. Two composite samples are collected, one during normal operations and one during cleaning. Some facilities may take more samples because of multiple outfalls.
- **Settleable Solids (SS)** – measured quarterly. Two composite samples are collected, one during normal operations and one during cleaning. Some facilities may take more samples because of multiple outfalls.
- **pH** – measured quarterly when settleable solids are measured.
- **Water Temperatures** – daily maximum and minimum water temperatures are measured within the hatchery. Temperature units are recorded for egg development in some hatcheries. Effluent and receiving stream temperatures are measured weekly from April to October.
- **Dissolved Oxygen (DO)** – measured only when conditions warrant (e.g., periods of low flows and high temperatures).
- **Air Temperatures** – maximum and minimum temperatures are recorded daily at some stations, but there are no special monitoring requirements.
- **Flow Logs** – changes in water flows through the hatchery ponds are recorded whenever flows are altered for hatchery management activities (i.e., ponding of fish, splitting of fish lots, fish releases, etc.).

Objective 6: Communicate effectively with other fish producers, managers and the public.

Coordination/Communication within ODFW

**Annual Fish Production Meetings:** ODFW conducts meetings throughout the state to set annual fish production goals for all public hatcheries in Oregon. These meetings involve the participation of ODFW research, management and fish culture staff as well as representatives from applicable federal agencies and tribes.

**Record Keeping:** The following records are kept at all ODFW hatcheries:
• Anadromous Adult Transaction Report – details the collection and disposition of all adult fish handled at the facility.
• Mark Recovery Report – details sex, fish length and tag information from all marked adult fish that are captured.
• Egg and Fry Report – records all egg and fry movements, treatments, etc.
• Monthly Ponded Report – updates hatchery operations from the previous month (i.e., current number of fish, size, transfers or releases, feed conversion, mortality, medication, etc.).
• Monthly Progress Report – document summarizing operational activities for the hatchery and all satellite facilities (e.g., fish culture, fish health, fish distribution, maintenance and safety).
• Fish Loss and Treatment Report – records disease problems and daily mortality.
• Fish Loss Report/Investigation – when 1,000 or more juveniles or 10 or more adult fish are accidentally lost in a single accident.
• Predator Mortality Report – documents any fish predators that may die at the hatchery facility.
• Fish Liberation Reports – details information regarding all fish releases (e.g., fish numbers, size, location, method of release, marks, etc.).
• Coded –Wire Tag Release Reports – record of all juvenile fish released with coded-wire tags.
• Length Frequency Record – details fish lengths of all anadromous fish released (based on a sample of the releases).
• Chemical use, waste discharge monitoring, purchasing, budget, hazardous materials, safety, vehicles, equipment, maintenance and alarm logs.
• Visitor Log – some facilities record the daily visitor use of the facility; however, this is not a requirement.

**Hatchery Management System (HMS):** Computerized system to collect, report, summarize and analyze hatchery production data. This system is a tool to be used in production control at all hatchery management levels.

**Interagency Coordination Communication**

Pacific Northwest Fish Health Protection Committee (PNFHPC): This group is comprised of representatives from U.S. and Canadian fish management agencies, tribes, universities, and private fish operations. The groups meets twice a year to monitor regional fish health policies and to discuss current fish health issues in the Pacific Northwest.

In-Season Communications: Communication with PAC, takes place each year to coordinate proper fish and egg transfers in an effort to meet basin-wide goals at all facilities, where applicable.

**Communication with the General Public**

Salmon River Hatchery receives approximately 2,800 visitors each year, and an average of 12 tours are given in a year.
Salmon River Hatchery
Fall Chinook Salmon – Stock 36H (Salmon River)

Salmon River Hatchery
260,000 Green Eggs
October

Release

Salmon River
200,000 @ 14 fpp
August – September
Salmon River Hatchery
Coho Salmon – Stock 13H (Big Creek)

Salmon River Hatchery
410,000 Eyed Eggs
from Big Creek Hatchery
December

S Fk Klaskanine (CCF)
385,000 @ 30 fpp
October

S Fork Klaskanine River
385,000 @ 15 fpp
April
Salmon River Hatchery
Rainbow Trout – Stock 72T (Cape Cod Triploid)

Salmon River Hatchery
40,000 @ 75 fpp
from Roaring River Hatchery
June

7 Waterbodies – North Coast District
14,300 @ 2 fpp
March

4 Waterbodies – Mid Coast District
15,700 @ 2 fpp, 4,500 @ 1 fpp
March – June